

Gene therapy for arthritis

Review Article

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Summary

Gene therapy for the treatment of arthritis is developing rapidly. The ability to deliver genes to local sites of inflammation decreases the possibility of systemic side effects, making arthritis a good candidate for gene therapy. Animal models of arthritis provide a means of testing gene transfer strategies. Several issues still need to be addressed including which genes to deliver, how to deliver these genes, and how to regulate gene expression in vivo.

I. Introduction

Arthritis is a major health problem among working age people in the United States, with greater than 2 million men and greater than 3 million women reporting activity limitation (Yelin, 1992). The most prevalent form of arthritis, rheumatoid arthritis (RA), affects approximately 0.8% of most populations (Koopman, 1997). Arthritic symptoms have been reported for 55% of individuals 70 years of age. Among these elderly affected individuals, 3/4 were limited in physical actions and 1/3 were limited in daily living (Yelin, 1992). Therefore, the chronic symptoms of arthritis impact significantly on the quality of life.

Until recently, the treatment of arthritis, particularly RA, involved the use of non-specific anti-inflammatory agents, such as nonsteroidal anti-inflammatory agents (NSAIDs), steroids and methotrexate. These anti-rheumatic drugs allow relief of many symptoms of the disease, but can exhibit harmful side effects and do not necessarily alter the natural course of the disease (Koopman, 1997). The cause of RA is unknown, and the mechanism of action of many of the drugs used to treat RA remains unknown. However, recent studies of molecular and cellular mechanisms that govern the pathophysiology of arthritis has led to the discovery of therapeutic biological agents that offer greater specificity in the treatment of arthritis. These biological agents are currently being delivered primarily at the protein level. The

short half-life of these molecules necessitates frequent re-administration. These naturally-produced molecules have the potential to be delivered via gene transfer, which may allow for a reduction in the requirement for frequent re-administration of the drug.

II. Gene delivery strategies

A. In vivo gene transfer

Although RA often affects local joints, immunological responses observed in patients with RA demonstrate the presence of systemic components of arthritis. Therefore, the treatment of RA can be approached with either systemic or local therapies.

Systemic gene delivery, such as by i.v. administration, has been demonstrated in animal models. These studies have examined mainly the short-term effects on arthritis and not the long-term systemic effects, including potential toxicity. Therefore, the delivery of genes directly into the bloodstream requires further investigation.

For the treatment of arthritis, local gene delivery is an attractive therapeutic option. Since the target of arthritis is the synovium or cells contained within the affected joint, local therapies involving injection directly into the affected joint space, could potentially provide delivery of genes to a limited space and reduce toxic systemic effects. Local

injection of adenovirus encoding a reporter gene to inflamed joints of monkeys with CIA results in expression that is contained to the synovium and is not present in other tissue samples, indicating that gene transfer to synovial tissue may be safe in primates and may exhibit an ideal biodistribution (Goossens, 1997). However, local administration of adenovirus in other animal models has effects on distal joints, suggesting that local delivery of gene products may produce systemic effects that must be analyzed appropriately (Bakker, 1997; Ghivizzani, 1998; May, 1998).

B. Ex vivo gene transfer

The ex vivo approach involves removal of synovial cells, culturing and infection of these cells with the appropriate virus, usually retrovirus, and subsequently returning the cells to the joint space. This procedure, while cumbersome and expensive, also provides for analysis and selection of the genetically altered cells before returning them to the joint space.

III. Gene transfer vectors

A. Viral vectors

Various gene therapy vectors have been utilized that can be grouped mainly into viral and non viral vectors. Since viruses naturally deliver genetic material to cells, the use of viruses is the basis for most gene delivery systems. Viruses are the most widely used means of delivering genes in arthritic animal models (Nita, 1996). Among viral vectors, retroviral and adenoviral vectors are primarily used for gene delivery, and both have particular characteristics that make them suitable for the delivery of genes in the treatment of arthritis.

Adenoviruses are easily produced at high titres and infect non dividing synovial cells. Adenovirus delivery of the β -galactosidase gene intra-articularly demonstrates that adenovirus can infect non dividing synovial cells and β -galactosidase expression can last up to 21 days (Sawchuck, 1996). Intravenous administration of adenovirus encoding vIL-10 also indicates that vIL-10 can be detected up to 7 days after injection and can inhibit CIA (Apparailly, 1998; Ma, 1998). However, adenoviral vectors induce an inflammatory response, which may come from the viral proteins being expressed or the transgene product itself. In terms of gene expression, adenoviral encoded proteins are normally short-lived, which is thought to be due to this inflammatory process.

Retroviruses are produced at relatively low titres, infect only dividing cells, and incorporate into the host genome. Retroviral vectors are primarily used ex vivo to transfect cultured synovial cells that divide, allowing for retroviral infection. Recent studies indicate that stimulation of cells

with TNF in vitro allows retroviral transduction of cells (Jorgensen, 1997), and that inflamed synovium, which produces TNF, may be more susceptible to retroviral uptake (Ghivizzani, 1997). These findings suggest that retroviral vectors might be delivered intra-articularly to target inflamed synovium. Long term gene expression is desirable for any gene therapy vector. Incorporation of the retrovirus into the host genome allows for long term gene expression; however, with this incorporation the risk of insertional mutagenesis exists. Unlike adenovirus-infected cells, retrovirus-infected cells have not been a target for destruction by the immune system (Evans, 1997).

Another kind of viral vector, the lentivirus, is derived from retroviruses, but has the capability to infect non dividing cells (Naldini, 1996). This virus may have promise for targeting non dividing synovial cells in the treatment of arthritis.

B. Non viral vectors

Various methods of non viral gene delivery include liposomal delivery, direct plasmid injection, and gene gun delivery. Non viral DNA delivery offers low toxicity, but most methods available are very inefficient at transfection of synovial cells. Gene transfer to rabbit and rat synovial cells by direct plasmid injection demonstrates that plasmid uptake resembles non specific endocytosis (Yovandich, 1995). The transient expression of the reporter plasmid corresponds with the degradation of plasmid DNA, indicating that intra-articular injection of plasmid DNA results in short-term gene expression. Long term gene expression of non viral plasmid DNA vectors has been achieved in muscle tissue (Tripathy, 1996). Expression of certain genes in skeletal muscle via plasmid injection has systemic effects on the immune system (Raz, 1993). Plasmid DNA encoding TGF- β delivered into thigh muscle of rats with streptococcal cell wall induced arthritis, suppressed the chronic disease and virtually eliminated subsequent inflammation and arthritis (Song, 1998). Therefore, intra-muscular injection of plasmid may be a less toxic way to systemically deliver anti-inflammatory products for the treatment of arthritis.

IV. Candidate genes

Analysis of cytokine expression between arthritic and non-arthritic joints indicates an increase in a number of cytokines in arthritic joints. This information has led to two main strategies to reduce inflammation in arthritic joints. The first approach involves the use of natural inhibitors of pro-inflammatory cytokines. The second approach, immune deviation, involves administration of cytokines that naturally down regulate pro-inflammatory cytokine synthesis.

A. Natural inhibitors of inflammatory cytokines

TNF- and IL-1 are major regulators of inflammation in arthritic joints. Inhibitors of these two cytokines reduce arthritis in both animal models of arthritis and in ongoing human trials. In collagen-induced arthritis (CIA), an animal model of RA, treatment with antibody to TNF (Joosten, 1994; Thorbecke, 1992) or IL-1 (Geiger, 1993; Joosten, 1994; Joosten, 1996; Thorbecke, 1992) reduced disease severity. In human trials administration of cA2, an antibody specific for TNF, dramatically suppressed symptoms of disease, although this effect required continual treatment (Elliott, 1994; Elliott, 1994; Elliott, 1993).

1. Interleukin-1 receptor antagonist (IL-1Ra)

IL-1Ra regulates IL-1 activity in vivo by binding to IL-1 receptors. IL-1Ra, while inhibiting IL-1 from binding, itself does not stimulate activity through the IL-1 receptor. However, a 10-100 fold excess of IL-1Ra over IL-1 is necessary to block the effects of IL-1 activity in vivo (Dinarello, 1991; Hirsch, 1996). Continuous administration of high levels of IL-1Ra can block CIA (Joosten, 1996; Wooley, 1993). Transgenic mice overproducing IL-1Ra exhibit a reduction in the incidence and severity of CIA, and mice lacking IL-1Ra have a significantly earlier onset of CIA (Ma, 1998). Human trials aimed at determining the efficacy of administration of recombinant human IL-1Ra are still being assessed (Campion, 1996). These studies indicate that IL-1Ra is a good candidate gene for reduction of arthritis.

Several animal models of arthritis have shown benefits after IL-1Ra gene delivery. Expression of human IL-1Ra in rabbits with antigen induced arthritis changed the course of arthritis and suppressed the effects of IL-1 (Bandara, 1993; Otani, 1996). Ex vivo retroviral transduction of primary synoviocytes grafted to ankle joints in rats with bacterial cell wall-induced arthritis showed a significantly suppressed severity of recurrence of arthritis and attenuated erosion of cartilage and bone (Makarov, 1996). Treatment of mice with CIA by ex vivo transduction of NIH/3T3 fibroblasts with retrovirus expressing human IL-1Ra prevented CIA in injected knee joints and the "draining" paws (Bakker, 1997). Rabbits treated with adenovirus expressing human IL-1Ra had both in vitro and in vivo effects, including inhibition of IL-1 activity and inhibition of induced prostaglandin E2 synthesis. Therefore, IL-1Ra shows great promise as a gene to deliver for the treatment of arthritis.

Results from many of the above studies using IL-1Ra led the way to the first human gene therapy trials for RA which began in 1996. Using an ex vivo approach, cells removed from patients joints are transfected with retroviral vectors expressing IL-1Ra (McCarthy, 1996 and reviewed in Evan, 1998). The cells are tested for both IL-1Ra expression and for the presence of endotoxin and other agents. IL-

1Ra-transduced and untransduced cells are injected back into the joints, and removed at the time of joint replacement to determine whether expression of IL-1Ra was achieved. This human trial is the first step toward assessment of local gene therapy for RA.

2. Soluble TNF receptor (sTNFR)

sTNFR is a natural inhibitor of TNF activity. Two receptors for TNF have been isolated, p55 and p75, that bind both TNF and TNF (Loetscher, 1990; Smith, 1990). Soluble forms of these receptors, which are extracellular and contain ligand binding domains, inhibit TNF activity (Mohler, 1993). The sTNFR administered in clinical trials of RA is comprised of the soluble portion of the p75 cell surface receptor fused to the Fc portion of human IgG1 (sTNFRFc). The IgG1 portion prolongs the half-life of the molecule (Mohler, 1993). sTNFRFc inhibits both CIA (Mori, 1996; Williams, 1995; Wooley, 1993) and can dramatically suppress the arthritic symptoms of RA, although again, continuous administration is required (Moreland, 1997). Recently, sTNFR marketed under the trade name Enbrel (Immunex Corporation) has received approval by the FDA for the treatment of RA as a subcutaneous injection administered twice weekly.

Gene delivery of sTNFR in animal models has inhibitory effects on arthritis. In rats with CIA, systemic delivery of an adenoviral vector encoding sTNFR prior to or following the onset of arthritis, suppressed CIA. However, intra-articular administration of this vector induced an adenoviral synovitis, which was not overcome even by the expression of the sTNFR (Le, 1997). The transfer of CIA to SCID mice can also be inhibited by transducing DBA/1 spleen cells with retrovirus encoding sTNFR (Chernajovsky, 1995).

In other gene delivery studies, Ghivizzani, et al., injected adenoviruses encoding either IL-1Ra or sTNFR, both separately and in combination, into rabbit's knees (Ghivizzani, 1998). IL-1Ra reduced cartilage matrix degradation and white blood cell infiltration into the joint space. sTNFR by itself was not as effective as IL-1Ra. However, treatment with both IL-1Ra and sTNFR showed greater inhibition of white blood cell infiltration and cartilage breakdown with a considerable reduction in synovitis. In addition, with both reagents, effects on contralateral control knees were also observed, suggesting that local intra-articular treatment may be used to treat systemic polyarticular arthritides.

B. Immune deviation

An imbalance between the activities of Th1 and Th2 cells is thought to play a role in the pathophysiology of many autoimmune diseases, such as RA. Th1 cells secrete cytokines such as IL-2 and IFN-, that normally mediate

pro-inflammatory immune responses, whereas Th2 cells secrete cytokines such as IL-4, IL-10, and IL-13 that can downregulate Th1 activity.

Administration of IL-4, IL-10 and IL-13 proteins to CIA mice indicate that these cytokines can inhibit the disease process (Bessis, 1996; Hesse, 1996; Horsfall, 1997; Joosten, 1997; Tanaka, 1996; Walmsley, 1996). Another animal model using streptococcal cell wall fragments to induce arthritis in rats, also demonstrates that IL-4 administration can reduce pro-inflammatory cytokine production and can inhibit experimental arthritis (Allen, 1993). In human RA synovial cells, IL-4 and IL-10 also have inhibitory effects on pro-inflammatory cytokine production (Chomarat, 1995; Isomaki, 1996; Katsikas, 1994; van Roon, 1996). These studies indicate that the Th2 type cytokines IL-4, IL-10, IL-13, which can inhibit pro-inflammatory cytokine production and the arthritic process in animal models, are good candidates for gene transfer.

1. Viral IL-10 (vIL-10) gene therapy

vIL-10 is homologous to both mouse and human IL-10 and shares many of their immunosuppressive properties, but lacks their immunostimulatory properties (Go, 1990; MacNeil, 1990). Systemic administration of adenovirus encoding vIL-10 before the onset of CIA inhibited arthritis (Apparailly, 1998; Ma, 1998), but the effects were short-term, probably due to the inflammatory response to the adenovirus. Local administration of vIL-10 in the footpad (Whalen, 1998) or intra-articularly into the knee (Ma, 1998) reduced the incidence of arthritis, indicating again that local gene expression can have systemic effects on disease.

2. Fas Ligand

Other methods that eliminate proliferating synovial cells are also being investigated, even though the removal of synovium has not been a successful cure for arthritis. The transduction of synovial cells with adenovirus that expresses Fas ligand induced apoptosis of synovial cells producing Fas. Administration of the virus into inflamed joints ameliorated CIA in DBA/1 mice (Zhang, 1997).

V. Future gene therapy for arthritis

Much progress has been made in recent years in the field of gene therapy for arthritis. Future efforts will be focused on determining which genes are the most promising for therapy, which vectors are the best for delivering these genes, and ultimately how to regulate expression of the genes being delivered.

References

- Allen JB, Wong HL, Costa GL, Bienkowski MJ, and Wahl SM (1993). Suppression of monocyte function and differential regulation of IL-1 and IL-1ra by IL-4 contribute to resolution of experimental arthritis. **J Immunol** 151, 4344-4351.
- Apparailly F, Verwaerde C, Jacquet C, Auriault C, Sany J, and Jorgensen C (1998). Adenovirus-mediated transfer of viral IL-10 gene inhibits murine collagen-induced arthritis. **J Immunol**. 160, 5213-5220.
- Bakker AC, Joosten LA, Arntz OJ, Helsen MM, Bendele AM, van de Loo FAJ, and van den Berg WB (1997). Prevention of murine collagen-induced arthritis in the knee and ipsilateral paw by local expression of human interleukin-1 receptor antagonist protein in the knee. **Arthritis Rheum** 40, 893-900.
- Bandara G, Mueller GM, Galea-Lauri J, Tindal MH, Georgescu HI, Suchanek MK, Hung GL, Glorioso JC, Robbins PD, and Evans CH (1993). Intra-articular expression of biologically active interleukin-1-receptor antagonist protein by *ex vivo* transfer. **Proc. Natl. Acad. Sci.** 90, 10764-10768.
- Bessis N, Boissier MC, Ferrara P, Blankenstein T, Fradelizi D, and Fournier C (1996). Attenuation of collagen-induced arthritis in mice by treatment with vector cells engineered to secrete interleukin-13. **J Immunol**. 26, 2399-2403.
- Campion GV, Lebsack ME, Lookabaugh J, Gordon G, and Catalano M (1996). Dose-range and dose-frequency study of recombinant human interleukin-1 receptor antagonist in patients with rheumatoid arthritis. The IL-1Ra Arthritis Study Group. **Arthritis Rheum** 39, 1092-1101.
- Chernajovsky Y, Adams G, Podhajcer OL, Mueller GM, Robbins PD, and Feldmann M (1995). Inhibition of transfer of collagen-induced arthritis into SCID mice by *ex vivo* infection of spleen cells with retroviruses expressing soluble tumor necrosis factor receptor. **Gene Ther** 2, 731-735.
- Chomarat P, Banchereau J, and Miossec P (1995). Differential effects of interleukins 10 and 4 on the production of interleukin-6 by blood and synovium monocytes in rheumatoid arthritis. **Arthritis Rheum**. 38, 1046-1054.
- Dinarello CA (1991). Interleukin-1 and interleukin-1 antagonism. **Blood** 77, 1627-1652.
- Elliott MJ, Maini RN, Feldmann M, Kalden JR, Antoni C, Smolen JS, Leeb B, Breedveld FC, Macfarlane JD, Bijl H, and et al. (1994). Randomised double-blind comparison of chimeric monoclonal antibody to tumour necrosis factor alpha (cA2) versus placebo in rheumatoid arthritis. **Lancet** 344, 1105-1110.
- Elliott MJ, Maini RN, Feldmann M, Long-Fox A, Charles P, Bijl H, and Woody JN (1994). Repeated therapy with monoclonal antibody to tumour necrosis factor alpha (cA2) in patients with rheumatoid arthritis. **Lancet** 344, 1125-1127.
- Elliott MJ, Maini RN, Feldmann M, Long-Fox A, Charles P, Katsikis P, Brennan FM, Walker J, Bijl H, Ghayeb J, and et al. (1993). Treatment of rheumatoid arthritis with chimeric

- monoclonal antibodies to tumor necrosis factor alpha. **Arthritis Rheum** 36, 1681-1690.
- Evans CH, and Robbins PD (1997). Getting genes into human synovium [editorial; comment]. **J Rheumatol** 24, 2061-2063.
- Evans CH, Whalen JD, Ghivizzani SC, and Robbins PD (1998). Gene therapy in autoimmune diseases. **Ann Rheum Dis** 57, 125-127.
- Geiger T, Towbin H, Cosenti-Vargas A, Zingel O, Arnold J, Rordorf C, Glatt M, and Vosbeck K (1993). Neutralization of interleukin-1 beta activity *in vivo* with a monoclonal antibody alleviates collagen-induced arthritis in DBA/1 mice and prevents the associated acute-phase response. **Clin. Exp. Rheu.** 11, 515-522.
- Ghivizzani SC, Lechman ER, Kang R, Tio C, Kolls J, Evans CH, and Robbins PD (1998). Direct adenovirus-mediated gene transfer of interleukin 1 and tumor necrosis factor alpha soluble receptors to rabbit knees with experimental arthritis has local and distal anti-arthritic effects. **Proc Natl Acad Sci USA** 95, 4613-4618.
- Ghivizzani SC, Lechman ER, Tio C, Mule KM, Chada S, McCormack JE, Evans CH, and Robbins PD (1997). Direct retrovirus-mediated gene transfer to the synovium of the rabbit knee: implications for arthritis gene therapy. **Gene Ther** 4, 977-982.
- Go NF, Castle BE, Barrett R, Kastelein R, Dang W, Mosmann TR, Moore KW, and Howard M (1990). Interleukin 10, a novel B cell stimulatory factor: unresponsiveness of X chromosome-linked immunodeficiency B cells. **J Exp Med** 172, 1625-1631.
- Goossens P, Bout B, t'Hart B, Brok H, Breedveld FC, Valerio D, and Huizinga T (1997). Possibility and safety of gene transfer to inflamed synovial tissue after intra-articular administration. **Arthritis Rheum.** S221.
- Hesse M, Bayrak S, and Mitchison A (1996). Protective major histocompatibility complex genes and the role of interleukin-4 in collagen-induced arthritis. **Eur J Immunol** 26, 3234-3237.
- Hirsch E, Irikura VM, Paul SM, and Hirsh D (1996). Functions of interleukin 1 receptor antagonist in gene knockout and overproducing mice. **Proc Natl Acad Sci USA** 93, 11008-11013.
- Horsfall AC, Butler DM, Marinova L, Warden PJ, Williams RO, Maini RN, and Feldmann M (1997). Suppression of collagen-induced arthritis by continuous administration of IL-4. **J. Immunol.** 159, 5687-5696.
- Isomaki P, Lukkainen R, Saario R, Toivanen P, and J. P (1996). Interleukin-10 functions as an antiinflammatory cytokine in rheumatoid synovium. **Arthritis Rheum.** 39, 386-395.
- Joosten LAB, Helsen MMA, van de Loo FAJ, and van den Berg WB (1994). Amelioration of established collagen-induced arthritis (CIA) with anti-IL-1. **Agents Actions Special Conference Issue** 41, C174-C176.
- Joosten LAB, Helsen MMA, van de Loo FAJ, and van den Berg WB (1996). Anticytokine treatment of established type II collagen-induced arthritis in DBA/1 mice. **Arthritis Rheum.** 39, 797-809.
- Joosten LAB, Lubberts E, Durez P, Helsen MMA, Jacobs MJM, Goldman M, and van den Berg WB (1997). Role of interleukin-4 and interleukin-10 in murine collagen-induced arthritis. **Arthritis Rheum.** 40, 249-260.
- Jorgensen C, Demoly P, Noel D, Mathieu M, Piechaczyc M, Gougat C, Bousquet J, and Sany J (1997). Gene transfer to human rheumatoid synovial tissue engrafted in SCID mice [see comments]. **J Rheumatol** 24, 2076-2079.
- Katsikas PD, Chu C-Q, Brennan FM, Maini RN, and Feldmann M (1994). Immunoregulatory role of interleukin 10 in rheumatoid arthritis. **J. Exp. Med.** 179, 1517-1527.
- Koopman W. (1997). Arthritis and Allied Conditions. A textbook of Rheumatology (Baltimore, MD: Williams and Wilkins).
- Le CH, Nicolson AG, Morales A, and Sewell KL (1997). Suppression of collagen-induced arthritis through adenovirus-mediated transfer of a modified tumor necrosis factor alpha receptor gene. **Arthritis Rheum** 40, 1662-1669.
- Loetscher H, Pan YC, Lahm HW, Gentz R, Brockhaus M, Tabuchi H, and Lesslauer W (1990). Molecular cloning and expression of the human 55 kd tumor necrosis factor receptor. **Cell** 61, 351-359.
- Ma Y, Thornton S, Boivin GP, Hirsh D, Hirsch R, and Hirsch E (1998). Altered susceptibility to collagen-induced arthritis in transgenic mice with aberrant expression of interleukin-1 receptor antagonist. **Arthritis Rheum.** 41, 1798-1805.
- Ma Y, Thornton S, Duwell LE, Bluestone JA, and Hirsch R (1998). Gene therapy with vIL-10 inhibits CIA. **J. Immunol.** 161, 1516-1524.
- MacNeil IA, Suda T, Moore KW, Mosmann TR, and Zlotnik A (1990). IL-10, a novel growth cofactor for mature and immature T cells. **J Immunol** 145, 4167-4173.
- Makarov SS, Olsen JC, Johnston WN, Anderle SK, Brown RR, Baldwin AS, Jr., Haskill JS, and Schwab JH (1996). Suppression of experimental arthritis by gene transfer of interleukin 1 receptor antagonist cDNA. **Proc Natl Acad Sci USA** 93, 402-406.
- McCarthy M (1996). Gene therapy for rheumatoid arthritis starts clinical trials. **Lancet** 348, 323.
- Mohler KM, Torrance DS, Smith CA, Goodwin RG, Stremmler KE, Fung VP, Madani H, and Widmer MB (1993). Soluble tumor necrosis factor (TNF) receptors are effective therapeutic agents in lethal endotoxemia and function simultaneously as both TNF carriers and TNF antagonists. **J Immunol** 151, 1548-1561.
- Moreland LW, Baumgartner SW, Schiff MH, Tindall EA, Fleischmann RM, Weaver AL, Ettlinger RE, Cohen S, Koopman WJ, Mohler K, Widmer MB, and Blosch CM (1997). Treatment of rheumatoid arthritis with a recombinant human tumor necrosis factor receptor (p75)-Fc fusion protein. **N Engl J Med** 337, 141-147.

- Mori L, Iselin S, DeLibero G, and Lesslauer W (1996). Attenuation of collagen-induced arthritis in 55-kDa TNF receptor type 1 (TNFR1)-IgG1-treated and TNFR1-deficient mice. **J. Immunol.** 157, 3178-3182.
- Naldini L, Blomer U, Gallay P, Ory D, Mulligan R, Gage FH, Verma IM, and Trono D (1996). In vivo gene delivery and stable transduction of nondividing cells by a lentiviral vector [see comments]. **Science** 272, 263-267.
- Nita I, Ghivizzani SC, Galea-Lauri J, Bandara G, Georgescu HI, Robbins PD, and Evans CH (1996). Direct gene delivery to synovium. **Arthritis Rheum.** 39, 820-828.
- Otani K, Nita I, Macaulay W, Georgescu HI, Robbins PD, and Evans CH (1996). Suppression of antigen-induced arthritis in rabbits by ex vivo gene therapy. **J. Immunol.** 156, 3358-3562.
- Raz E, Watanabe A, Baird SM, Eisenberg RA, Parr TB, Lotz M, Kipps TJ, and Carson DA (1993). Systemic immunological effects of cytokine genes injected into skeletal muscle. **Proc Natl Acad Sci USA** 90, 4523-4527.
- Sawchuck S, Boivin GP, Duwel LE, Ball W, Bove K, Trapnell B, and Hirsch R (1996). Anti T cell receptor monoclonal antibody prolongs transgene expression following adenovirus-mediated in vivo gene transfer to the mouse synovium. **Hum. Gene Ther.** 7, 499-506
- Smith CA, Davis T, Anderson D, Solam L, Beckmann MP, Jerzy R, Dower SK, Cosman D, and Goodwin RG (1990). A receptor for tumor necrosis factor defines an unusual family of cellular and viral proteins. **Science** 248, 1019-1023.
- Song XY, Gu M, Jin WW, Klinman DM, and Wahl SM (1998). Plasmid DNA encoding transforming growth factor-beta1 suppresses chronic disease in a streptococcal cell wall-induced arthritis model. **J Clin Invest** 101, 2615-2621.
- Tanaka Y, Otsuka T, Hotokebuchi T, Miyahara H, Nakashima H, Kuga S, Nemoto Y, Niino H, and Niho Y (1996). Effect of IL-10 on collagen-induced arthritis in mice. **Inflamm. Res.** 45, 283-288.
- Thorbecke GJ, Shah R, Leu CH, Kuruvilla AP, Hardison AM, and Palladino MA (1992). Involvement of endogenous tumor necrosis factor and transforming growth factor during induction of collagen type II arthritis in mice. **Proc. Natl. Acad. Sci.** 89, 7375-7379.
- Tripathy SK, Svensson EC, Black HB, Goldwasser E, Margalith M, Hobart PM, and Leiden JM (1996). Long-term expression of erythropoietin in the systemic circulation of mice after intramuscular injection of a plasmid DNA vector. **Proc Natl Acad Sci USA** 93, 10876-10880.
- van Roon JAG, van Roy LAM, Gmelig-Meyling FHJ, Lafeber FPJG, and Bijlsma JWJ (1996). Prevention and reversal of cartilage degradation in rheumatoid arthritis by interleukin-10 and interleukin-4. **Arthritis Rheum.** 39, 829-835.
- Walmsley M, Katsikis PD, Abney E, Parry S, Williams RO, Maini RN, and Feldmann M (1996). Interleukin-10 inhibition of the progression of established collagen-induced arthritis. **Arthritis Rheum.** 39, 495-503.
- Whalen J, Lechman E, Robbins P, and Evans C. (1998). Gene transfer of the viral IL-10 gene to the mouse footpad can prevent collagen-type II induced arthritis. In 44th Annual Meeting, Orthopaedic Research Society (New Orleans, Louisiana), pp. 308.
- Williams RO, Ghayeb J, Feldmann M, and Maini RN (1995). Successful therapy of collagen-induced arthritis with TNF receptor-IgG fusion protein and combination with anti-CD4. **Immunology** 84, 433-439.
- Wooley PH, Whalen JD, Chapman DL, Berger AE, Richard KA, Aspar DG, and Staite ND (1993). The effect of an interleukin-1 receptor antagonist protein on type II collagen-induced arthritis and antigen-induced arthritis in mice. **Arthritis Rheum.** 36, 1305-1314.
- Yelin E (1992). Arthritis. The cumulative impact of a common chronic condition. **Arthritis Rheum.** 35, 489-97.
- Yovandich J, O'Malley B, Jr., Sikes M, and Ledley FD (1995). Gene transfer to synovial cells by intra-articular administration of plasmid DNA. **Hum Gene Ther** 6, 603-610.
- Zhang H, Yang Y, Horton JL, Samoiloiva EB, Judge TA, Turka LA, Wilson JM, and Chen Y (1997). Amelioration of collagen-induced arthritis by CD95 (Apo-1/Fas)-ligand gene transfer. **J Clin Invest** 100, 1951-1957.